

# Effects of carrier polymers on chloramphenicol-loaded electrospun matrices intended for the treatment of infected wounds

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## Introduction

Antimicrobial drug-loaded electrospun nanofibrous dressings are of major interest as novel topical drug delivery systems for managing chronic wound infections. Electrospinning is a simple and versatile process by which polymer nanofibers can be produced using an electrostatically driven jet of polymer solution or polymer melt (**Fig. 1**). Electrospun nanofibers have many useful properties for wound care applications, including oxygen permeability, high porosity and surface-to-volume ratio that can promote haemostasis and absorb wound exudates. Morphology of electrospun nanofibers is similar to natural extracellular matrix in the skin that promotes cell adhesion, migration and proliferation [1,2].

## Aims

To develop antibacterial electrospun nanofiber mats for the treatment of infected wounds and understand the effect of different carrier polymers on the relevant physicochemical, biopharmaceutical and antibacterial properties.

## Materials and Methods

Model antibacterial drug – Chloramphenicol (CAM)

Electrospinning solutions:

- I. PCL 12.5% in chloroform and methanol (3:1 V/V)
- II. PCL 10%, PEO 2% in chloroform and methanol (3:1 V/V)

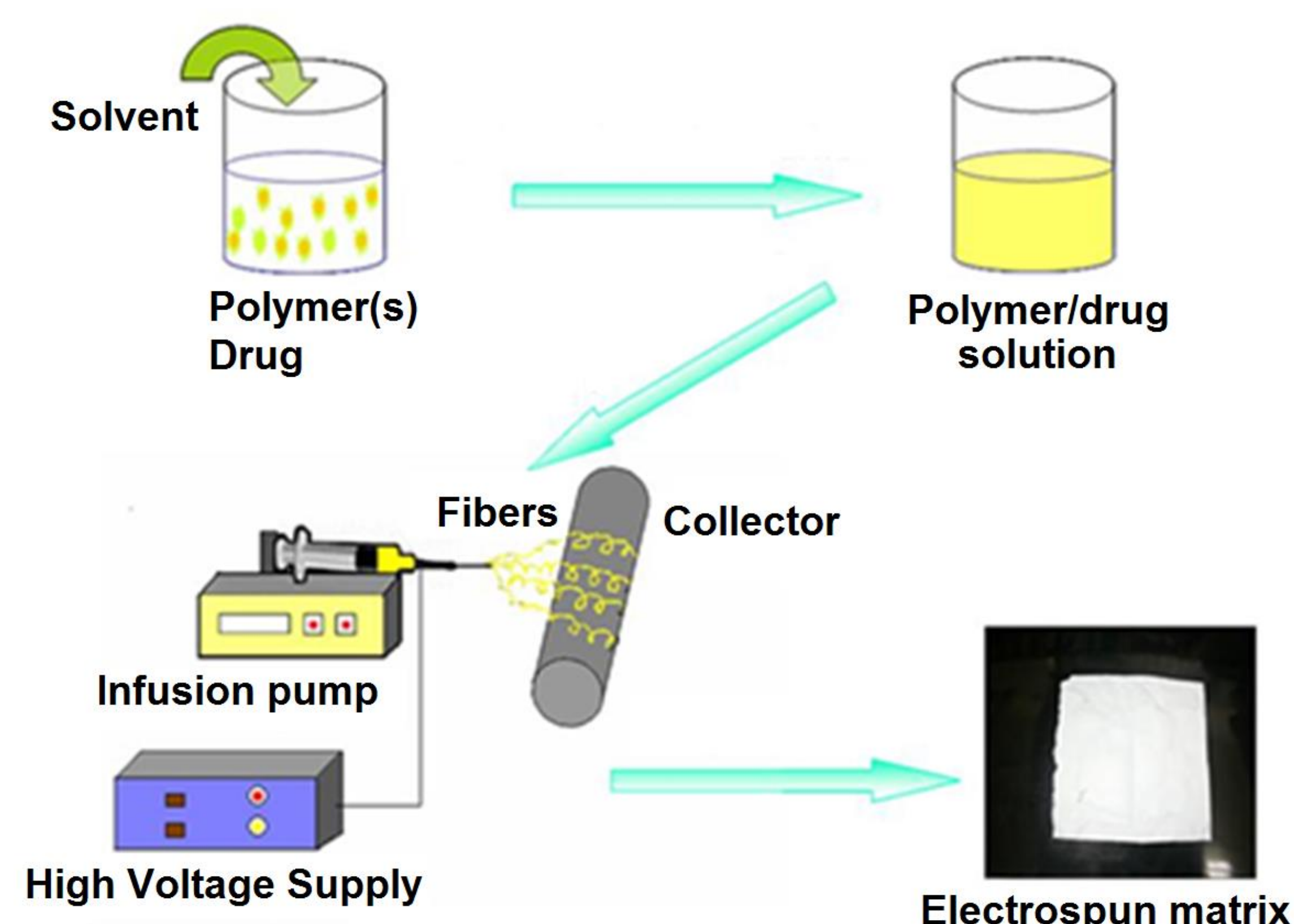
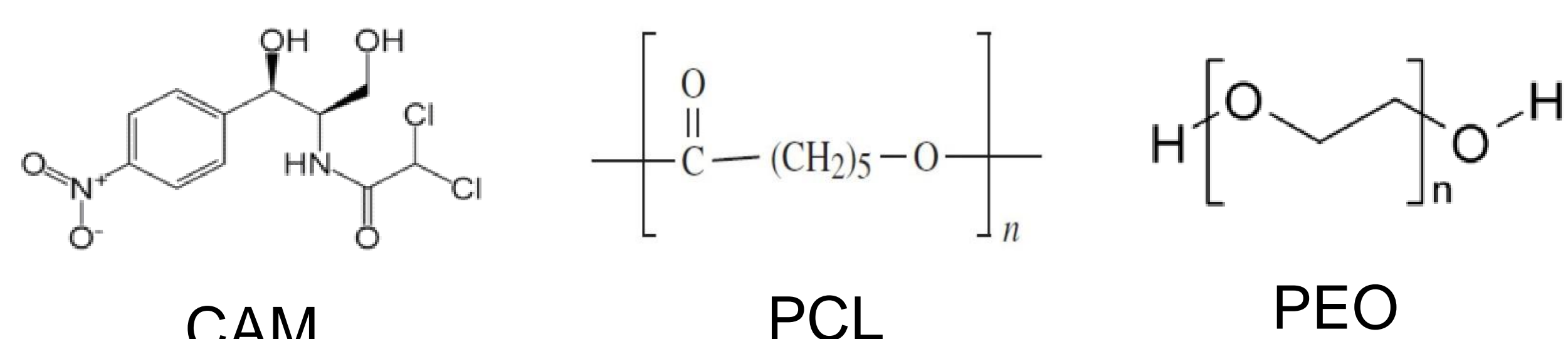


Figure 1: Experimental setup for preparing electrospun nanofibers [3].

Characterization of electrospun matrices:

- ❖ Physicochemical:
  - Scanning Electron Microscopy (SEM)
  - X-ray Powder Diffractometry (XRPD)
  - Differential Scanning Calorimetry (DSC)
- ❖ Biopharmaceutical:
  - Drug-release: Modified dissolution test
  - Antibacterial activity: Diffusion test on agar plates (*E. coli* MG1655, LB plates)

## Results

Morphology of the fibers

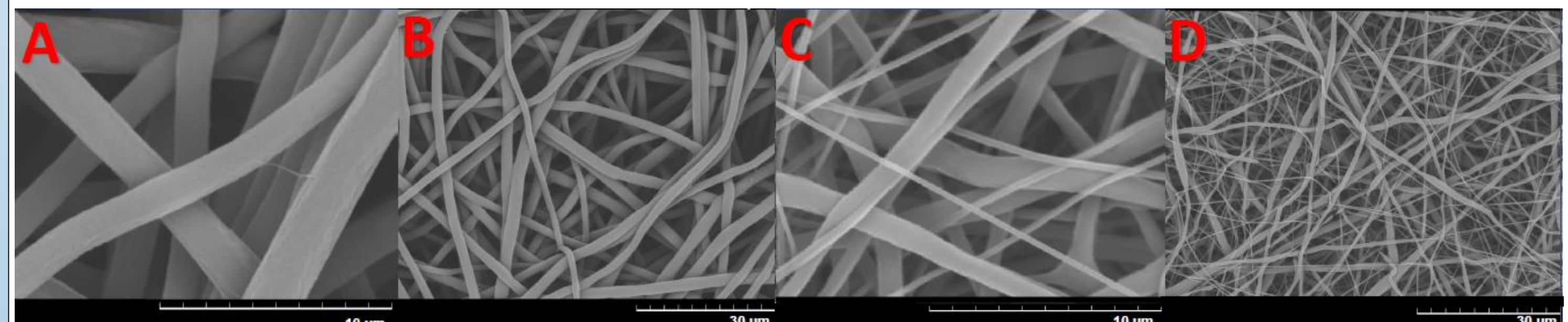


Figure 2: SEM micrographs of PCL + PEO + CAM 4% microfibers x10 000 (A), x2000 (B) and PCL + CAM 4% nanofibers x10 000 (C), x2000 (D).

Absence of characteristic diffraction reflections of CAM in the XRPD patterns of electrospun fibers suggests amorphous state of the drug (**Fig. 3**).

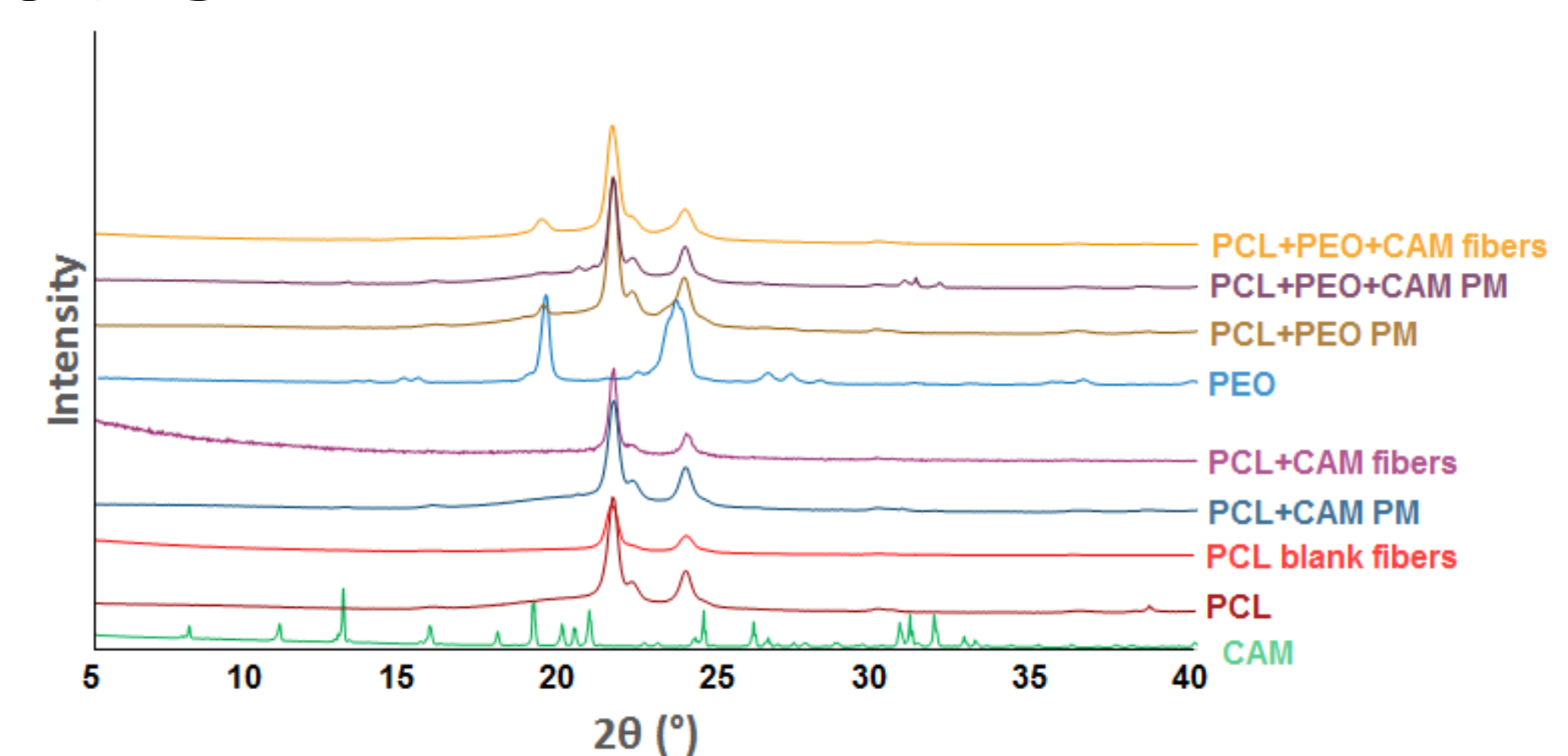


Figure 3: XRPD diffraction patterns of pure substances, physical mixtures and electrospun fibers.

Amorphous state of the drug in both fibers was further confirmed with DSC (**Figs. 4a and 4b**).

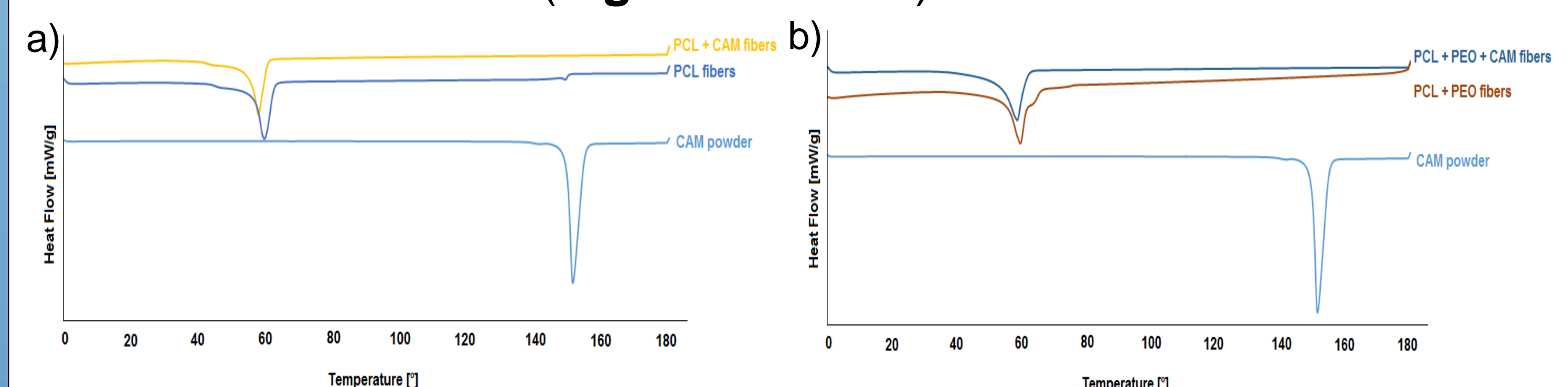


Figure 4: DSC thermograms of (a) PCL drug-loaded fibers and (b) PCL-PEO drug-loaded fibers, both with curves of blank fibers and CAM powder.

Dissolution studies showed that PCL fibers have initial burst release and following slow release of the drug, whereas PCL+PEO fibers release the drug remarkably faster (**Figs. 5a and 5b**).

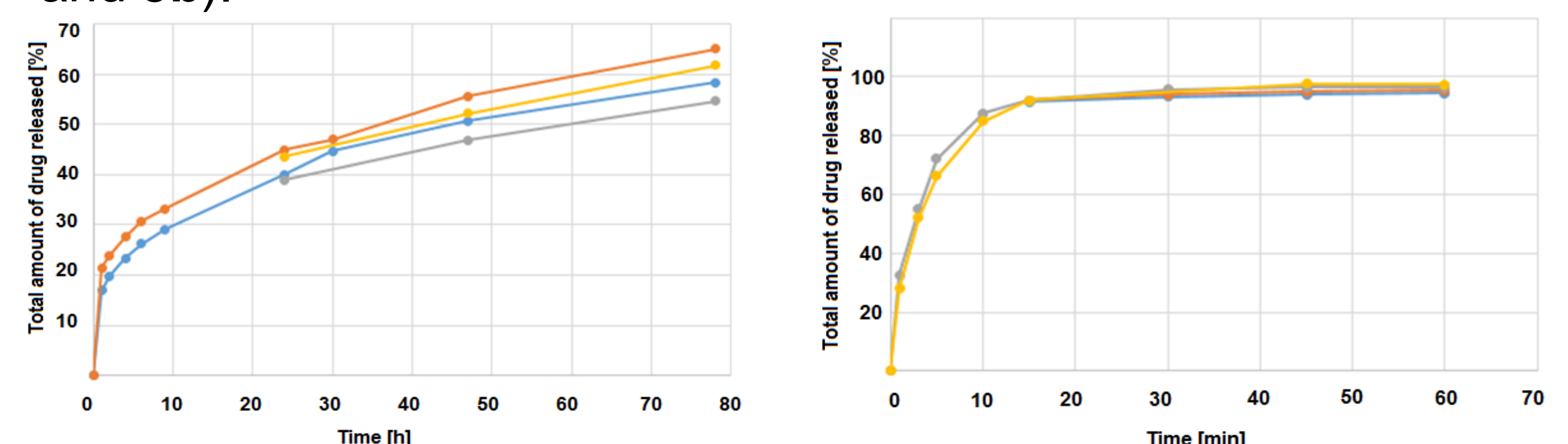


Figure 5: Drug release profiles of (a) PCL drug-loaded fibers and (b) PCL-PEO drug-loaded fibers.

Clear areas of inhibition formed around all drug-loaded fibers (**Fig. 6**).

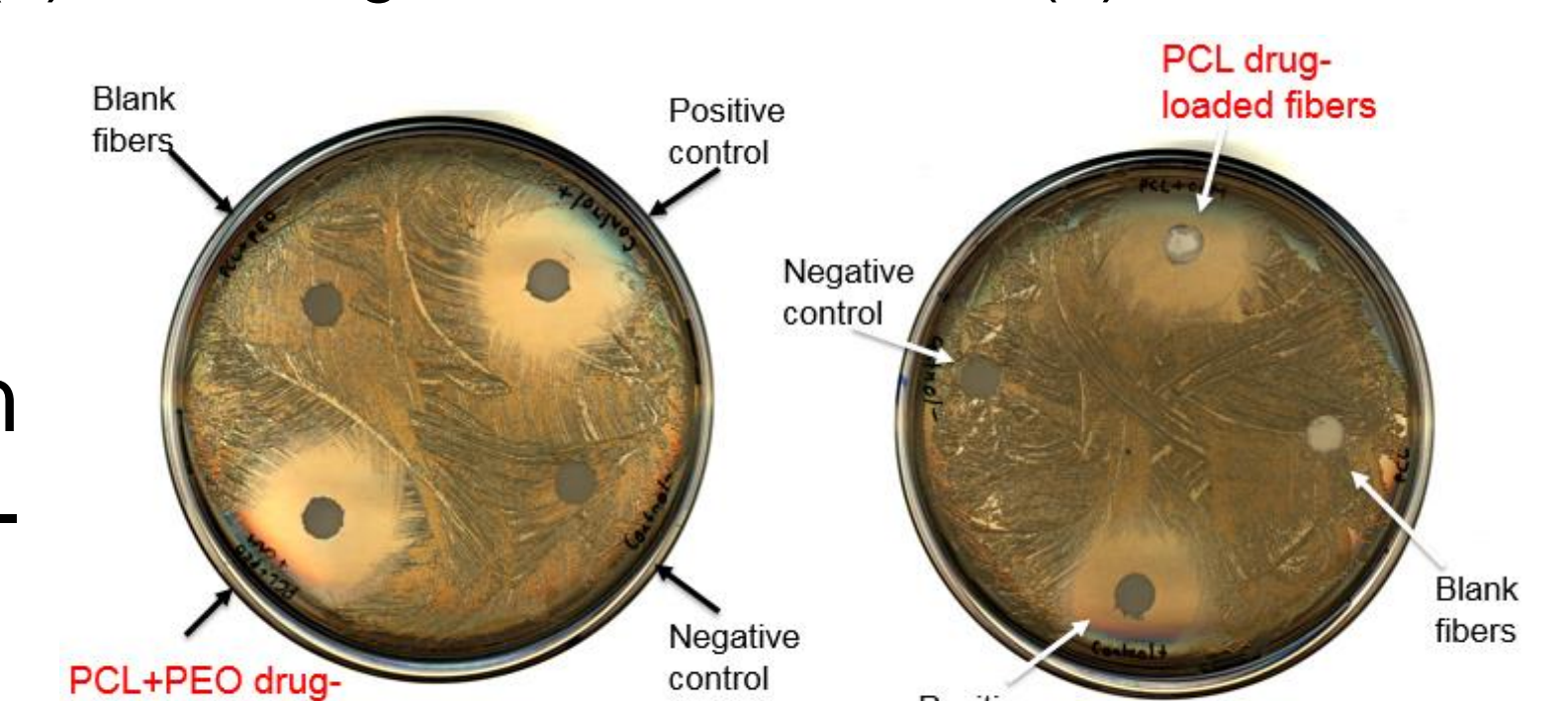


Figure 6: Disc diffusion test on agar plates.

The developed mats have great potential to be used for successful wound therapy.

## References

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2. Lee SY, Kuti JL, Nicolau DP. 2005. Antimicrobial management of complicated skin and skin structure infections in the era of emerging resistance. Surg. Infect. (Larchmt). 6:283–95.
- 3 Retrieved from: <http://faculty.ksu.edu.sa/waheed.almasry/Pages/NanofiberTechnology.aspx>